This article was downloaded by:

On: 27 January 2011

Access details: Access Details: Free Access

Publisher Taylor & Francis

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-

41 Mortimer Street, London W1T 3JH, UK



Nucleosides, Nucleotides and Nucleic Acids

Publication details, including instructions for authors and subscription information: http://www.informaworld.com/smpp/title~content=t713597286

A Convenient Approach to the Synthesis of Azido-Acyclic Nucleosides

Takashi Ogawa^a; Hiroshi Takaku^a; Naoki Yamamoto^b

^a Laboratory of Bioorganic Chemistry, Department of Industrial Chemistry, Chiba Institute of Technology, Narashino, Chiba, Japan ^b Departments of Virology and Parasitology, Yamaguchi University School of Medicine, Ube, Japan

To cite this Article Ogawa, Takashi , Takaku, Hiroshi and Yamamoto, Naoki(1989) 'A Convenient Approach to the Synthesis of Azido-Acyclic Nucleosides', Nucleosides, Nucleotides and Nucleic Acids, 8: 4, 499 - 504

To link to this Article: DOI: 10.1080/07328318908054192 URL: http://dx.doi.org/10.1080/07328318908054192

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: http://www.informaworld.com/terms-and-conditions-of-access.pdf

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

A CONVENIENT APPROACH TO THE SYNTHESIS OF AZIDO-ACYCLIC NUCLEOSIDES

Takashi Ogawa, Hiroshi Takaku*, and Naoki Yamamoto*

Laboratory of Bioorganic Chemistry, Department of Industrial Chemistry, Chiba Institute of Technology, Tsudanuma, Narashino, Chiba 275, Japan and *Department of Virology and Parasitology, Yamaguchi University School of Medicine, Ube 755, Japan

Abstract

The azidation of unprotected acyclic nucleosides (4) was carried out in a one-pot reaction by means of the reagent triphenylphosphine-carbon tetraiodide-sodium azide to give the corresponding mono-azido-acyclic nucleosides (6) in good yields without by-products such as the di-azido-acyclic nucleosides.

INTRODUCTION

A number of acyclic nucleoside analogues have been reported to be active against herpe simples virus. 1) Recently, we have also reported that some acyclic nucleoside cyclic phosphoroamidate derivatives showed inhibitory effects on the proliferation of tumor cells. 2) On the other hand, 3'-azido-2',3'-dideoxythymidine (AZT) is a potent and selective inhibitor of the replication of HIV, the human immunodeficiency virus responsible for ATDS. 3,4)

We now wish to report a convenient approach for the synthesis of azido-acyclic nucleoside derivatives ($\underline{6}$), which were evaluated for their anti-HIV activity.

CHEMISTRY

First, we examined the synthesis of acyclic nucleosides $(\underline{4a})$ using the procedure of Ogilvie without the removal of benzyl groups of acyclic sugar moiety through catalytic hydrogenation (Scheme 1).⁵⁾ The chloromethyl derivative $(\underline{1})$ was treated with trimethylsilylated thymine in the presence of n-Bu₄NI to give the corresponding product $\underline{2}$ in 42% yield. Compound $\underline{2}$ thus obtained was treated with potassium acetate in DMF to give the acetoxy derivative $\underline{3}$ which was then treated with methanolic

ammonia to remove the acetyl groups. 1-[[2-Hydroxy-1-(hydroxy-methyl)ethoxy]methyl]thymine ($\underline{4a}$) was obtained in 10% yield after separation by silica gel chromatography. Compared with the route using compound $\underline{5}$ as a starting material, it is noted that yields of the desired acyclic nucleosides were undoubtedly low.

Next, we examined the direct conversion of unprotected acyclic nucleosides $(\underline{4})$ to azido-acyclic nucleosides $(\underline{6})$ using the procedure of Hata et al.⁶). Hata and coworkers reported⁶) that the most effective combination was triphenylphosphine-carbon tetrabromide-lithium azide, which gave a good yield of 5'-azido-deoxythymidine. In this method, when sodium azide was used in place of lithium azide, $1-\{(2-azido-1-(hydroxymethyl)-ethoxy\} methyl\}thymine <math>(\underline{6a})$ was obtained in an unsatisfactory yield $(12 \ \$)$. More recently, Ogilvie et al.⁷) reported that compound $\underline{4c}$ was treated with a combination of triphenylphosphine-carbon tetrabromide-lithium azide to give the desired product $\underline{6c}$ with

Scheme 2

9-[[2-azido-1-(azidomethyl)ethoxy]methyl]adenine as a by-product. However, we tried the reaction of <u>4a</u> with triphenylphosphine-carbon tetraiodide-sodium azide in dry DMF at room temperature for 24 h; the corresponding azido-product (<u>6a</u>) was obtained in a good yield (75%) without by-products such as 1-[[2-azido-1-(azidomethyl)ethoxy]methyl]thymine. In a similar manner, the other azido-acyclic nucleosides (<u>6b,c</u>) were obtained in <u>66%</u> and 68% yields, respectively. In these reactions, by-products such as cyclonucleosides were not detected at all.

Antiviral Activity

These compounds $(\underline{6})$ have been—evaluated for citotoxicity and inhibition of HIV replication in MT4 cells, but no activities were detected.

EXPERIMENTAL

All melting points were determined on a Yanagimoto micromelting point apparatus (hot stage type) and are uncorrected. Ultraviolet spectra were recorded on a Shimazu UV-200 spectrophotometer. The $^1\text{H-NMR}$ spectra were recorded on a JEOL JNM-MH 100 spectrometer in CDCl $_3$ with trimethylsilane as an internal standard. Thin-layer chromatography (t.l.c) was performed on plates of Kiesegel 60F_{254} (Merck). Column chromatography was performed on silica gel (BW-300; Fuji Davison Co. Ltd.).

1,3-Dichloro-2-chloromethoxypropane (1). This preparation is carried out by a modification of the procedure of Ogilvie et al.⁵⁾. 1,3-Dichloro-2-propanol (14.8 ml, 155 mmol) was dis-

solved in $\mathrm{CH_2Cl_2}$ (50 ml). Paraformaldehyde (9.8 g) was added and the mixture was cooled in an ice-salt bath. Dry HCl gas was bubbled into the solution with stirring for 8 h. Anhydrous $\mathrm{CaCl_2}$ was added to the solution and the reaction mixture was filtered through Celite. The filtrate was evaporated in vacuo to give 24.8 g (90%) of compound 1 as a syrup. $^1\mathrm{H-NMR}$ (CDCl₃) & 3.75 (d, 4H, $\mathrm{CH_2-CH-CH_2}$), 4.07 (m, 1H, $\mathrm{CH_2-CH-CH_2}$), 5.59 (s, 2H, $\mathrm{CH_2Cl}$).

The product was used directly in coupling experiments without further purification.

1-[[2-Chloro-1-(chloromethyl)ethoxy]methyl]thymidine (2). Thymine (3.5 g, 27.7 mmol) was suspended in HMDS (20 ml) and ammonium sulfate (0.12 g, 0.9 mmol) was added. The mixture was heated under reflux temperature until a clear solution was obtained. The excess HMDS was removed under reduced pressure and the residue was dissolved in dry CH_2Cl_2 (30 ml) and $\underline{1}$ (12.3 g, 70 mmol) and $(n-Bu)_4NI$ (0.31 g, 0.83 mmol) were added. solution was stirred at room temperature for 12 h. was quenched with addition of a mixture of MeOH and H2O (4:1, v/v) and the solution was evaporated in vacuo. The residue was dissolved in CH2Cl2 (50 ml), washed with water(50 ml X 3), and dried over Na2SO4. The solvent was concentrated to a small volume (10 ml) and the solution was applied to a silica gel column which was eluted with a stepwise gradient of MeOH (0-10%) in The fractions containing the desired product were CH₂Cl₂. pooled and the solvent was evaporated in vacuo to give compound 3a (3.1 g, 42%) as a gum; UV λ max(MeOH) 264 nm; 1 H-NMR (CDCl₃) δ 1.91 (s, 3H, CH₃), 3.60 (s, 4H, \underline{CH}_2 -CH- \underline{CH}_2), 3.90 (m, 1H, $CH_2-CH-CH_2$), 5.20 (s, 2H, CH_2-N), 7.60 (s, 1H, H-6). Anal. Calc. for C9H12N2O3Cl2: C, 40,47; H, 4,53; N, 10,49. Found: C, 40,28; H, 4,31; N, 10.35.

Conversion of 2 to 4a. Compound 2 (1.87 g, 11.5 mmol) was mixed with potassium acetate (4.53 g, 46 mmol) in dry DMF (30 ml) and the reaction mixture was heated at reflux for 3 h under The mixture was filtered through Celite. nitrogen atmosphere. The filtrate was evaporated in vacuo and the residue was treated with methanolic ammonia for 6 h. The solution was concentrated under reduced pressure and the residue was applied to a silica gel column which was eluted with CH_2Cl_2 -MeOH (9:1, v/v). The fractions containing the desired product were collected and the solvent was evaporated in vacuo. The residue was crystallized from MeOH to give compound 4a (270 mg, 10%); mp 159-160°C (lit⁸) mp 155-156 °C); UV \times max(H₂O, pH 7.0) 264; ¹H-NMR (DMSO-

 d_6) & 1.73 (s, 3H, CH_3); 3.47 (d, 4H, $\underline{CH_2}$ -CH- $\underline{CH_2}$); 4.01 (m, 1H, $\underline{CH_2}$ - \underline{CH} - \underline{CH} - \underline{CH}); 5.18 (s, 2H, \underline{CH} - \underline{N}); 7.57 (s, 1H, H-6).

Direct conversion of 4 to 6. To a mixture of acyclic nucleosides (2.0 mmol), triphenylphosphine (1.05 g, 4.0 mmol), and sodium azide (1.3 g, 4.0 mmol) in dry DMF (40 ml) was added carbon tetraiodide (2.0 g, 4.0 mmol). The mixture was stirred at room temperature for 24 h, MeOH (1 ml) was added, the solvent was removed in vacuo, and chromatography on a column of silica gel [eluted with a stepwise gradient of MeOH (0-10%) in CH_2Cl_2] gave mono-azido derivatives 6. Yields were 6a, 75%; 6b, 66%, and 6c, 68%.

6b: mp: 213-215 °C; UV \nearrow max(MeOH) 265 nm; IR (KBr) ? 2091 cm⁻¹ (N₃); ¹H-NMR (DMSO-d₆) \$ 3.60 (m, 5H, CH₂-CH-CH₂), 4.21 (m, 1H, OH), 5.35 (s, 2H, CH₂N), 5.82 (d, 1H, J_{5,6}=7.5 Hz, H-5), 7.69 (d, 1H, J_{5,6}=7.5 Hz, H-6). Anal. Calcd for C₈H₁₃N₆O₂: C, 42,66; H, 5.82; N, 37,32. Found: C, 42,39; H, 5.94; N, 37.41.

6c: mp 198-200°C (lit⁷⁾ mp: 200-204°C); UV \uparrow max(MeOH) 259 nm; IR (KBr) \uparrow 2100 cm⁻¹ (N₃); ¹H-NMR (DMSO-d₆) & 3.57 (m, 5H, CH₂-CH-CH₂), 4.00 (m, 1H, OH), 5.79 (s, 2H, CH₂N), 7.25 (br s, 2H, NH₂), 8.21 and 8.31 (s, 2H, H-2 and H-8). Anal. Calcd for C₉H₁₂N₈O: C, 43,54; H, 4,87; N, 45,14. Found: C, 43,61; H, 4,98; N, 45.25.

Acknowledgments

We thank Yodogawa Pharmaceutical Co. Ltd. for the gift of thymine, adenine, and cytosine.

REFERENCES

- For reviews, see R.J. Remy and J.A. Secrist III, Nucleosides Nucleotides, 4, 411 (1985).
- H. Takaku, T. Ito, S. Yoshida, T. Aoki, and E. De Clercq, Nucleosides Nucleotides, 6, 793 (1987).
- H. Mitsuya, K.J. Weinhold, P.A. Furman, M.H. St. Clair, S.N. Lehrman, R.C. Gallo, D. Bolognesi, D.W. Barry, and S. Broder, Proc.Natl.Acad.Sci.USA, 82, 7096 1985).
- R. Yarchoan, R.W. Klecker, K.J. Weihold, P.D. Markham, H.K. Lyerly, D.T. Durack, E. Gelmann, S.N. Lehrman, R.M. Blum, D.W. Barry, G.M. Shearer, M.A. Fischl, H. Mitsuya, R.C. Gallo, J.M.

- Collins, D.P. Bolognesi, C.E. Myers, S. Broder, Lancet, 1, 575 (1986); H. Mitsuya and S. Broder, Proc.Ntal.Acad.Sci.USA, 83, 1911(1986).
- 5. K.K. Ogilvie and D.M. Dixit, Nucleosides Nucleotides, $\underline{2}$, 147(1983).
- I. Yamamoto, M. Sekine, and T. Hata, J.Chem.Soc.Perkin Trans.
 306(1980).
- 7. K.K. Ogilvie, N. Nguyen-ba, M.F. Gillen, B.K. Radatus, U.O. Cheriyan, K.O. Smith, and K.S. Galloway, Can.J.Chem., <u>62</u>, 241 (1984).
- 8. K.K. Ogilvie, R.G. Hamilton, M.F. Gillen, and B.K. Radatus, Can.J.Chem., 62, 16(1984).

Received September 15, 1988.